ABSTRACT

"Mosquito malaria vectors cloak their legs to resist

insecticides"

Mary Kefi^{1,2}, Vasileia Balabanidou², Jason Charamis ^{1,2,} ,Panagiotis Ioannidis², Victoria Ingham ^{3,4}, Hilary Ranson³, John Vontas ^{2,5}

¹Department of Biology, University of Crete, Vassilika Vouton, 71409, Heraklion, Greece

²Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology-Hellas, 73100 Heraklion, Greece

³Department of Vector Biology, Liverpool School of Tropical Medicine, Pembroke Place, Liverpool, UK

⁴Parasitology Unit, Universitätsklinikum Heidelberg, Im Neuenheimer Feld 324, 69120 Heidelberg, Germany

⁵Pesticide Science Laboratory, Department of Crop Science, Agricultural University of Athens, 11855 Athens, Greece

ABSTRACT

While malaria continues to claim more than 400,000 lives each year (WHO, 2019), insecticide resistance is rapidly increasing in spectrum and intensity across Africa [1]. *Anopheles* legs are the key-tissues for insecticide uptake, as they comprise the first barrier the insecticide has to bypass to reach its neuronal targets [2]. In this context we use high-throughput methods to shed light to resistant mechanisms present in the first defense line of malaria vectors.

Our data on *An. coluzzii* leg-specific transcriptome provides valuable insights into the legs of pyrethroid resistant and short-term deltamethrin-exposed mosquitoes. The results suggest that xenobiotic detoxification is likely occurring in legs, while the enrichment of sensory proteins, ABCG transporters and cuticular genes is also evident. Constitutive resistance is primarily associated with elevated levels of detoxification and cuticular genes, while short-term insecticide-induced tolerance is linked with overexpression of transporters, GPCRs and GPCR-related genes, sensory/binding and salivary gland proteins [3].

Additionally, comparative proteomic analysis of the legs from resistant and susceptible mosquitoes revealed that resistant mosquitoes thicken their leg cuticles via enhanced

deposition of cuticular proteins, chitin filaments and, remarkably, cuticular hydrocarbons (CHCs) [4].

The last decarbonylation step of CHCs biosynthesis is catalyzed by CYP4Gs in oenocytes. We then focused on these enzymes and using *Drosophila melanogaster* as a tool we indicated the distinct contribution of each enzymes to CHC biosynthesis [5].

Structural and functional alterations in *Anopheles* legs are associated with reduced insecticide penetration that intensifies and potentially broadens resistance phenotype, and might affect other major physiological functions as well.

REFERENCES

- 1. Cook J, Tomlinson S, Kleinschmidt I, Donnelly MJ, Akogbeto M, Adechoubou A, Massougbodji A, Okê-Sopoh M, Corbel V, Cornelie S *et al*: **Implications of insecticide resistance for malaria vector control with long-lasting insecticidal nets: trends in pyrethroid resistance during a WHO-coordinated multi-country prospective study**. *Parasites & Vectors* 2018, **11**(1):550.
- Andriessen R, Snetselaar J, Suer RA, Osinga AJ, Deschietere J, Lyimo IN, Mnyone LL, Brooke BD, Ranson H, Knols BG: Electrostatic coating enhances bioavailability of insecticides and breaks pyrethroid resistance in mosquitoes. Proceedings of the National Academy of Sciences 2015, 112(39):12081-12086.
- Kefi M, Charamis J, Balabanidou V, Ioannidis P, Ranson H, Ingham VA, Vontas J: Transcriptomic analysis of resistance and short-term induction response to pyrethroids, in Anopheles coluzzii legs. 2021.
- Balabanidou V, Kefi M, Aivaliotis M, Koidou V, Girotti JR, Mijailovsky SJ, Juárez MP, Papadogiorgaki E, Chalepakis G, Kampouraki A: Mosquitoes cloak their legs to resist insecticides. Proceedings of the Royal Society B 2019, 286(1907):20191091.
- Kefi M, Balabanidou V, Douris V, Lycett G, Feyereisen R, Vontas J: Two functionally distinct CYP4G genes of Anopheles gambiae contribute to cuticular hydrocarbon biosynthesis. Insect biochemistry and molecular biology 2019, 110:52-59.